

Cyclodipeptide synthase-dependent pathways: an efficient tool to incorporate non-canonical amino acids into 2,5-diketopiperazines

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Cyclodipeptide synthases (CDPSs) constitute a family of bacterial enzymes that divert aminoacyl-tRNAs (AA-tRNAs) from the ribosomal machinery to synthesize 2,5-diketopiperazines (2,5-DKPs)^[1], a class of natural products containing many therapeutically promising compounds^[2].

CDPSs utilize two AA-tRNAs as substrates for catalysing formation of two peptide bonds between aminoacyl moieties to yield the DKP scaffold^[1], which is often equipped with different chemical groups by 2,5-DKP-tailoring enzymes found associated to CDPSs in dedicated biosynthetic pathways (Figure 1)^[3,4].

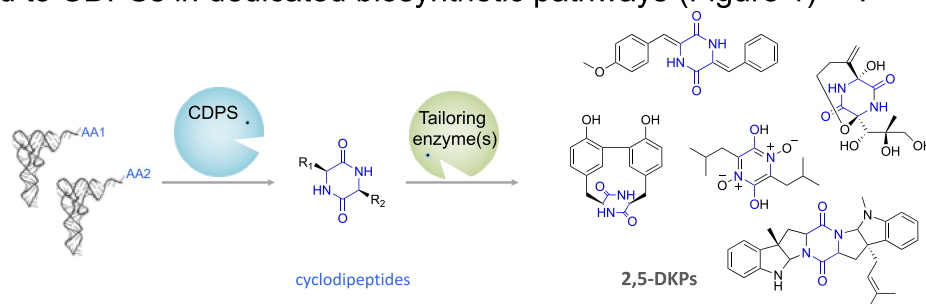


Figure 1: CDPS-dependent pathways

The recent characterization of large sets of CDPSs has revealed that they can produce highly diverse products^[5,6], and therefore have great potential for use in the production of different 2,5-DKPs. Several strategies for further increasing the diversity accessible with these enzymes will be discussed, including the incorporation of non-canonical amino acids by CDPSs^[7] and use of the remarkable diversity of 2,5 DKP-tailoring enzymes discovered in recent years^[8–10].

References:

- [1] M. Gondry, L. Sauguet, P. Belin, R. Thai, R. Amouroux, C. Tellier, K. Tuphile, M. Jacquet, S. Braud, M. Courçon, et al., *Nat Chem Biol* **2009**, *5*, 414–420.
- [2] A. D. Borthwick, *Chem Rev* **2012**, *112*, 3641–3716.
- [3] P. Belin, M. Moutiez, S. Lautru, J. Seguin, J. L. Pernodet, M. Gondry, *Nat Prod Rep* **2012**, *29*, 961–979.
- [4] M. Moutiez, P. Belin, M. Gondry, *Chem Rev* **2017**, *117*, 5578–5618.
- [5] I. B. Jacques, M. Moutiez, J. Witwinowski, E. Darbon, C. Martel, J. Seguin, E. Favry, R. Thai, A. Lecoq, S. Dubois, et al., *Nat Chem Biol* **2015**, *11*, 721–727.
- [6] M. Gondry, I. B. Jacques, R. Thai, M. Babin, N. Canu, J. Seguin, P. Belin, J. L. Pernodet, M. Moutiez, *Front Microbiol* **2018**, *9*, 46.
- [7] N. Canu, P. Belin, R. Thai, I. Correia, O. Lequin, J. Seguin, M. Moutiez, M. Gondry, *Angew. Chem. Int. Ed. Engl.* **2018**, *57*, 3118–3122.
- [8] P. Dubois, I. Correia, F. Le Chevalier, S. Dubois, I. Jacques, N. Canu, M. Moutiez, R. Thai, M. Gondry, O. Lequin, et al., *Sci. Rep.* **2019**, *9*, 9208.
- [9] P. Borgman, R. D. Lopez, A. L. Lane, *Org. Biomol. Chem.* **2019**, *17*, 2305–2314.
- [10] N. Canu, M. Moutiez, P. Belin, M. Gondry, *Nat. Prod. Rep.* **2019**, to be published.